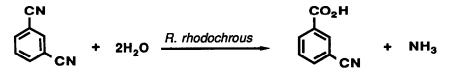
BACTERIA IN ORGANIC SYNTHESIS: SELECTIVE CONVERSION OF 1,3-DICYANOBENZENE INTO 3-CYANOBENZOIC ACID

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Summary: Suspensions of <u>Rhodococcus</u> <u>rhodochrous</u> NCIB 11,216 catalyse hydrolysis of dinitriles into cyanocarboxylic acids under mild conditions. This bioconversion was used for the highly selective synthesis of 3cyanobenzoic acid from 1,3-dicyanobenzene.

Acid catalysed hydrolysis of nitriles usually requires prolonged boiling with a concentrated solution of acid and therefore cannot be applied to acid sensitive or epimerisable molecules. Although recently milder catalytical procedures for nitrile hydrolysis were developed,¹ they are complex and require onerous isolation and purification steps. Furthermore, by chemical means, it is virtually impossible to selectively convert one cyanogroup of a polynitrile. Biological hydrolysis of nitriles² offers an interesting alternative as it can be performed at moderate temperatures and at pH values close to neutrality. Several researchers reported bacterial hydrolysis of aliphatic³ and aromatic⁴ nitriles into corresponding carboxylic acids or amides, however the processes are not adapted to selective conversion of polynitriles and very little is apparently known about the applicability of such a technique for practical synthetic purposes.

We observed that resting cells of <u>Rhodococcus</u> <u>rhodochrous</u> NCIB 11,216, which were previously grown on benzonitrile, selectively hydrolysed aromatic dinitriles into correpsonding cyanocarboxylic acids. We now report the first results as applied to the preparation of 3-cyanobenzoic acid from 1,3dicyanobenzene in nearly quantitative yield.



The following experiment is representative: Cells of <u>R. rhodochrous</u> (maintained on 1.5% w/v agar, containing per litre 10 g triptone, 5 g yeast extract and 5 g NaCl), were grown on the pH 7 basal salts medium containing per litre: 6 g Na₂HPO₄, 3 g KH₂PO₄, 0.5 g NaCl, 0.2 g MgSO₄. 7H₂O and 5 ml trace metals solution⁵ with 10 mM benzonitrile as the sole carbon and nitrogen source. Cultures were grown aerobically at 28 ^oC in 2 1 flasks containing 750

ml medium on a reciprocal shaker at 120 rpm and bacterial growth was monitored by measuring the optical density at 690 nm. Cells were harvested by centrifugation after 20 hr, in the early-mid exponential stage (1.2-1.7 OD) and washed with 0.1 M phosphate buffer at pH 7. The cells (2 g wet weight) were then resuspended in 0.25 M phosphate buffer pH 7 and incubated on a shaker with 0.57 g (4.45 mmol) of 1,3-dicyanobenzene at 30 $^{\circ}$ C for 25 hr. Aliquots were withdrawn periodically and reaction progress was monitored by quantitative determination of ammonia released into solution⁶. Upon release of 1 eq. of ammonia the reaction came to a standstill. Since the cells were shown to be active when incubated with a fresh batch of the dinitrile, evidently the bioconversion stopped at the stage of monohydrolysis. Acidification of the supernatant followed by extraction with chloroform afforded pure 3-cyanobenzoic acid in 95% yield⁷.

The kinetics of ammonia release indicated that the benzonitrile grown cells are similarly effective in half hydrolysis of 1,4-dicyanobenzene to form the corresponding 4-cyanobenzoic acid. However, when these cells were incubated with aliphatic dinitriles such as: $NC-(CH_2)_n-CN$ (n=3,4,5), hydrolysis proceeded at a very low rate and was not selective. On the other hand, resting cells of the bacterium, which were previously grown on propionitrile, had a remarkably different substrate affinity, and hydrolysed effectively (although not selectively) the above mentioned aliphatic dinitriles rather than the aromatic.

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